

Wnt/ β -Catenin Signaling Mediates the Antitumor Activity of Magnolol in Colorectal Cancer Cells^[S]

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ABSTRACT

Abnormal activation of the canonical Wnt/ β -catenin pathway and up-regulation of the β -catenin/T-cell factor (TCF) response to transcriptional signaling play a critical role early in colorectal carcinogenesis. Therefore, Wnt/ β -catenin signaling is considered an attractive target for cancer chemotherapeutic or chemopreventive agents. Small molecules derived from the natural products were used in our cell-based reporter gene assay to identify potential inhibitors of Wnt/ β -catenin signaling. Magnolol, a neolignan from the cortex of *Magnolia obovata*, was identified as a promising candidate because it effectively inhibited β -catenin/TCF reporter gene (TOPflash) activity. Magnolol also suppressed Wnt3a-induced β -catenin translocation and subsequent target gene expression in human embryonic kidney 293 cells. To further investigate the precise mechanisms of action in the regulation of Wnt/ β -catenin signaling by magnolol, we performed Western blot analysis, real-time reverse tran-

scriptase-polymerase chain reactions, and an electrophoretic mobility shift assay in human colon cancer cells with aberrantly activated Wnt/ β -catenin signaling. Magnolol inhibited the nuclear translocation of β -catenin and significantly suppressed the binding of β -catenin/TCF complexes onto their specific DNA-binding sites in the nucleus. These events led to the down-regulation of β -catenin/TCF-targeted downstream genes such as *c-myc*, matrix metalloproteinase-7, and urokinase-type plasminogen activator in SW480 and HCT116 human colon cancer cells. In addition, magnolol inhibited the invasion and motility of tumor cells and exhibited antitumor activity in a xenograft nude mouse model bearing HCT116 cells. These findings suggest that the growth inhibition of magnolol against human colon cancer cells can be partly attributed to the regulation of the Wnt/ β -catenin signaling pathway.

Introduction

Colorectal cancer is one of the leading causes of cancer-related human morbidity and mortality worldwide. Although surgery is the most effective treatment for advanced colon cancer, recurrence frequently occurs within a few years after surgery. Therefore, it is logical to develop strategies based on the carcinogenic progression of common colorectal cancers to decrease the incidence of colon cancer. The dysregulation of the Wnt/ β -catenin signaling pathway has been considered to play an important role in colon carcinogenesis.

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The Wnt signaling pathway can be divided into two distinct pathways: a canonical signaling pathway mediated by β -catenin and a noncanonical signaling pathway regulated by Ca^{2+} or a small G protein such as Rho/Rac (Huelsken and Behrens, 2002). The control of β -catenin levels is a key feature of the canonical Wnt/ β -catenin signaling pathway. In the absence of Wnt activation, cytosolic β -catenin is constitutively phosphorylated through casein kinase I α and glycogen synthase kinase 3 β (GSK3 β), a component of the β -catenin destruction complex that contains axin and adenomatous polyposis coli (APC) as scaffolding proteins. The phosphorylated β -catenin is subsequently degraded by the β -TrCP-mediated ubiquitin-proteasome pathway (Ikeda et al., 1998; Liu et al., 2005). When Wnt proteins are present, they bind to the Frizzled (Fz) receptor and to low-density lipoprotein-related receptors 5 and 6. These events lead to the dissociation of β -catenin from the APC/axin/GSK3 β de-

ABBREVIATIONS: GSK3 β , glycogen synthase kinase 3 β ; APC, adenomatous polyposis coli; Fz, frizzled; TCF, T-cell factor; MMP, matrix metalloproteinase; uPA, urokinase-type plasminogen activator; HEK, human embryonic kidney; hFz1, human frizzled receptor type 1; PARP, poly(ADP-ribose) polymerase; DMEM, Dulbecco's modified Eagle's medium; FBS, fetal bovine serum; siRNA, small interfering RNA; PCR, polymerase chain reaction; CM, conditioned medium.

struction complex. Thus, unphosphorylated β -catenin accumulates in the cytoplasm and translocates into the nucleus, where β -catenin interacts with T-cell factor (TCF) and lymphoid-enhancing factor to activate the transcription of Wnt/ β -catenin-mediated target genes (Bienz, 2002; Cong and Varmus, 2004). The target genes include cell cycle-regulating genes (*c-myc* and cyclin D1) and genes related to metastasis and the invasion of cancer cells (MMP-7 and uPA) (He et al., 1998; Tetsu and McCormick, 1999).

Aberrant activation of Wnt signaling has been found in a variety of cancers including colon cancers (Bright-Thomas and Hargest, 2003; Phelps et al., 2009; Shan et al., 2009). In particular, APC mutations are the most prevalent genetic alterations in colorectal cancers and lead to the accumulation of β -catenin. They are also associated with the constitutive activation of the Wnt pathway. Indeed, mutations of APC (present in more than 80% of sporadic colon cancers) and/or β -catenin (present in approximately 10% of colon cancers) are found in colon cancers, suggesting that these mutations eventually activate the Wnt pathway in colorectal cancer cells. Thus, modulation of the Wnt/ β -catenin signaling pathway is an attractive candidate for developing a targeted therapy for colorectal cancers.

In the present study, we used a cell-based reporter gene assay to search for novel small molecule inhibitors of the Wnt signaling pathway. We identified magnolol as a potent inhibitor of the Wnt pathway. Magnolol also exhibits antiproliferative and antitumor activity in colon cancer cells with the modulation of the Wnt signaling pathway.

Magnolol is one of major components of the cortex of *Magnolia obovata* Thunb. (Magnoliaceae). *M. obovata*, a medicinal plant widely distributed in Japan, has been traditionally used for gastrointestinal disorders, constipation, and cough. Magnolol has also shown many biological activities, such as antimicrobial, antiasthmatic, and antiplatelet activities (Clark et al., 1981; Teng et al., 1988; Ko et al., 2003). In addition, antiproliferative and antimetastatic effects against various cancer cell lines, including colon cancer, hepatoma, leukemia, fibrosarcoma, melanoma, squamous carcinoma, thyroid carcinoma, and prostate cancer cells, have also been reported (Zhong et al., 2003; Battle et al., 2005; Ishitsuka et al., 2005; Kong et al., 2005; Huang et al., 2007; Lee et al., 2008). However, the precise mechanism of the inhibition of colon cancer cell growth remains to be identified. Herein, we provide evidence that magnolol may represent a new class of compounds that inhibit the Wnt pathway by inhibiting the transcriptional activity of β -catenin/TCF.

Materials and Methods

Cell Culture and Reagents. Human colorectal carcinoma (HCT116) cells, colorectal adenocarcinoma (SW480) cells, embryonic kidney (HEK293) cells, L cells, and L cells stably transfected with Wnt3a (L-Wnt3a) were obtained from the American Type Culture Collection (Manassas, VA). L cells are used to obtain control conditioned medium for comparison with Wnt3a conditioned medium from L-Wnt3a cells. Cells were grown in the media: Dulbecco's modified Eagle medium (DMEM) for HEK293 and L cells; DMEM with 100 μ g of hygromycin for HEK293-hFz1 cells; DMEM with 400 μ g of G418 for L-Wnt3a cells; and RPMI 1640 medium for HCT116 cells. These media were supplemented with 10% FBS and antibiotics-antimycotics [100 units/ml penicillin G sodium, 100 μ g/ml streptomycin, and 250 ng/ml amphotericin B (PSF)]. SW480 cells were grown in RPMI

1640 medium containing 25 mM HEPES, 10% FBS, and PSF. DMEM, RPMI 1640 medium, FBS, antibiotic-antimycotic solution, β -catenin-specific siRNA, and trypsin-EDTA were purchased from Invitrogen (Carlsbad, CA). Bovine serum albumin, trichloroacetic acid, TRI reagent, HEPES, mouse monoclonal anti- β -actin antibody, hygromycin B, and other agents unless otherwise indicated were purchased from Sigma-Aldrich (St. Louis, MO). Mouse monoclonal anti-*c-myc* was purchased from Santa Cruz Biotechnology, Inc. (Santa Cruz, CA). Mouse monoclonal anti-cyclin D1, anti- β -catenin, anti-PARP, and anti-GSK-3 β antibodies were purchased from BD Biosciences (San Diego, CA). Complete protease inhibitor cocktail was purchased from Roche Applied Science (Penzberg, Germany). Alexa Fluor 488 goat anti-mouse IgG and SYBR Gold staining solution were purchased from Invitrogen. Gene-specific primers for reverse transcriptase-polymerase chain reaction (PCR), real-time PCR, and scrambled siRNA were synthesized from Bioneer (Daejeon, Korea). AMV reverse transcriptase, dNTP mixture, oligo(dT)₁₅ primer, RNasin, Taq DNA polymerase, and the dual luciferase assay system were purchased from Promega (Madison, WI). The TCF-electrophoretic mobility shift assay kit was purchased from Panomics (Redwood City, CA).

Test Compound. Magnolol [4-allyl-2-(5-allyl-2-hydroxy-phenyl) phenol] (Fig. 1), isolated from *Magnolia obovata* (purity >98.5%), was provided by Dr. KiHwan Bae (Chungnam National University, Daejeon, Korea) and dissolved in 100% dimethyl sulfoxide.

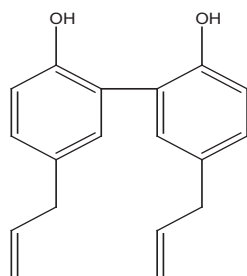
Reporter Gene Assay. Transient transfections were performed using Lipofectamine 2000 (Invitrogen). In brief, cells (1.5×10^4) were seeded onto a 48-well plate. TCF reporter plasmid contains two sets (with the second set in the reverse orientation) of three copies of the binding site (wild-type) upstream of the thymidine kinase minimal promoter and luciferase open reading frame (TOPflash, pGL3-OT). FOPflash (pGL3-OF) containing mutated TCF binding sites was used as a negative control. After 24 h, the cells were transfected with 0.1 μ g of the luciferase reporter constructs (TOPflash or FOPflash, respectively) and 0.005 μ g of the *Renilla reniformis* gene for normalization. HEK293 cells were also cotransfected with 0.02 μ g of pcDNA β -catenin and 0.008 μ g of the TCF4 plasmid. After 24 h of transfection, compound was added, and the cells were incubated for 24 h, lysed in $1 \times$ passive lysis buffer (Promega), and collected for luciferase and *R. reniformis* activity assays. Activity values were normalized to those for *R. reniformis* and are expressed as the relative value compared with the control.

Preparation of Wnt3a Conditioned Medium. Wnt3a conditioned medium (Wnt3a-CM) was prepared by culturing Wnt3a-secreting L cells (L-Wnt3a) in DMEM with 10% FBS for 4 days. The medium was harvested and sterilized using a 0.22- μ m filter. Fresh medium was added, and the cells were cultured for an additional 3 days. The medium was collected again and combined with the previous medium.

Stable Transfection. HEK293 cells (10^6 cells/dish in 100-mm dishes) were incubated for 24 h and transfected with the hFz1 plasmid (24 μ g; provided by Dr. S. Oh, Inje University, Gimhae, Korea). After 24 h, the cells were harvested and reseeded onto a 100-mm dish (10^4 cells/dish). Medium was replaced daily with fresh medium containing 200 μ g/ml hygromycin B until cell colonies were visualized. The cell colonies were then detached and reseeded onto a 24-well plate. To confirm stable transfection, a reporter gene assay was performed in the presence of Wnt3a-CM.

Isolation of Cytosolic and Nuclear Extracts. Cells (1×10^6 cells) were treated with test compounds for 24 h with or without Wnt3a-CM. Harvested cells were washed with PBS, suspended in ice-cold lysis buffer [10 mM Tris-HCl, pH 8.0, 1.5 mM MgCl₂, 10 mM KCl, 0.1 mM EDTA, 1 mM dithiothreitol, 2% NP-40, 50 mM sodium fluoride, 5 mM sodium orthovanadate, and protease inhibitor cocktail] on ice for 5 min. After centrifugation at 2500 rpm for 4 min at 4°C, the supernatant was collected as a cytosolic fraction, and the pellets were washed twice with ice-cold lysis buffer without NP-40. Cells were resuspended in hypertonic nuclear extract buffer [20 mM

A



B

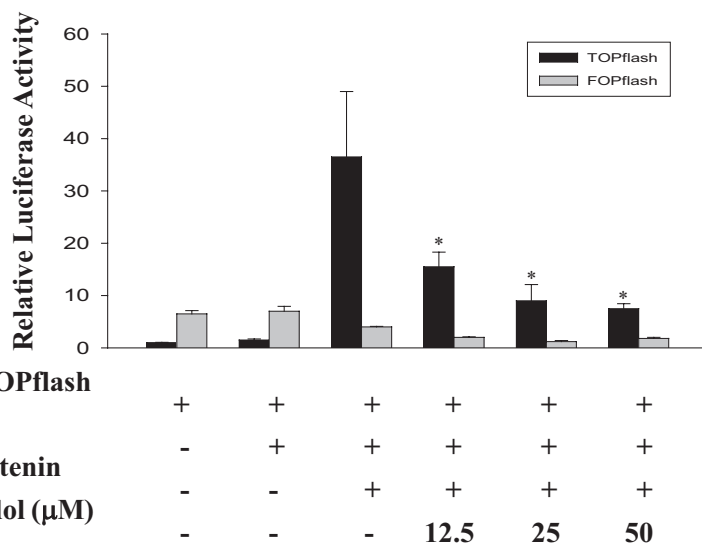


Fig. 1. Inhibition of β -catenin/TCF transcriptional activity by magnolol. A, chemical structure of magnolol. B, inhibitory effect of magnolol on TOPflash/FOPflash activity in HEK293 cells. HEK293 cells were transiently transfected with β -catenin and TCF4, TOPflash or FOPflash, and *R. reniformis* in the presence of magnolol. Results indicate the mean \pm S.D. ($n = 3$) and are representative of the findings from three or more separate experiments. *, $P < 0.05$, compared with the DMSO control.

Tris-HCl, pH 8.0, 420 mM NaCl, 1.5 mM $MgCl_2$, 0.2 mM EDTA, 25% glycerol, 50 mM sodium fluoride, and protease inhibitor cocktail on ice for 10 min and then centrifuged at 140,000 rpm for 15 min at 4°C. This supernatant, which contained nuclear extracts, was collected and stored in aliquots at -70°C . The protein content of the cell lysates was determined using the Bradford assay (Bradford, 1976).

Cell Proliferation Assay. Cells (5×10^3 cells/well in 96-well plates) were incubated for 24 h and treated with test compound for 24, 48, and 72 h. After incubation, cells were exposed to Premix WST-1 solution (10 μl /well; Takara, Shiga, Japan) for 2 to 4 h. Absorbance was measured at 450 nm. Cell viability was determined by comparing the absorbance of the vehicle-treated control group.

Western Blotting. Cells were treated with test compound for 24 h. Harvested cells were disrupted, and protein contents were measured by the Bradford assay. Equal amounts (40–80 μg) of protein samples were subjected to 6 to 15% SDS-polyacrylamide gel electrophoresis. Separated proteins were transferred to polyvinylidene difluoride membranes (Millipore Corporation, Billerica, MA). Membranes were incubated with primary antibodies diluted in 3% nonfat dry milk in phosphate-buffered saline-Tween 20 (1:200–1:2000) overnight at 4°C, washed three times with phosphate-buffered saline-Tween 20, and incubated with corresponding secondary antibodies. The blots were detected with an enhanced chemiluminescence detection kit (LabFrontier, Suwon, Korea) and a LAS 3000 Imager (Fuji Film Corp., Tokyo, Japan).

Reverse Transcription and Real-Time Polymerase Chain Reaction. HEK293 cells (4×10^5 cells/dish in 60-mm dishes) were transfected with 2 μg of pcDNA β -catenin for 24 h, and then cells were treated with test compound for 12 h. Total RNA was extracted using TRI reagent and reverse transcribed with AMV reverse transcriptase and the oligo(dT)₁₅ primer. PCR was performed in a mixture containing cDNA, 0.2 mM dNTP mixture, 10 pmol of gene-specific primers, and 0.25 unit of Taq DNA polymerase using a

GeneAmp PCR System 2400 (Applied Biosystems, Foster, CA). The PCR cycling parameters used are as follows: an initial denaturation step for 4 min at 94°C; 25 to 30 cycles of amplification, consisting of denaturation for 30 s at 94°C, annealing for 30 s at 55–57°C, and elongation for 30 s at 72°C; and a final extension step for 5 min at 72°C. The PCR products were separated by 2% agarose gel electrophoresis. The gel was stained with a SYBR Gold staining solution and visualized under a UV transilluminator (Alpha Imager; Alpha Innotech Corp., Santa Clara, CA). PCR primer sequences are listed in Supplemental Table 1.

Real-time PCR was conducted with a MiniOpticon system (Bio-Rad Laboratories, Hercules, CA), using 5 μl of reverse transcription product, iQ SYBR Green Supermix (Bio-Rad Laboratories), and primers for a total volume of 20 μl . The standard thermal cycler conditions were used: 95°C for 20 s, 40 cycles of 95°C for 20 s, 56°C for 20 s, and 72°C for 30 s, followed by 95°C for 1 min and 55°C for 1 min. The threshold cycle (C_T), the fractional cycle number at which the amount of amplified target gene reaches a fixed threshold, was determined by using MJ Opticon Monitor software. The mean threshold cycle (C_t) value for each transcript was normalized by dividing it by the mean C_t value for the β -actin transcript for that sample. Normalized transcript levels were expressed relative to sample obtained from the control. Real-time PCR primer sequences are listed in Supplemental Table 1.

Immunofluorescence. Cells were plated onto coverslips and incubated for 24 h. After incubation with test compound for 24 h, the cells were fixed with a 4% paraformaldehyde solution for 30 min at room temperature. The cells were then treated with a 0.1% Triton X-100 solution for permeabilization. After quenching with a 0.1% sodium borohydrate solution, the coverslips were blocked with blocking buffer (1% bovine serum albumin and 0.01% sodium azide in PBS) for 1 h. The coverslips were subsequently incubated with β -catenin antibodies diluted in blocking buffer (1:250) overnight at

4°C. The coverslips were further incubated with the corresponding fluorescence (Alexa Fluor 488)-labeled secondary antibodies for 1 to 2 h. The coverslips were washed three times with PBS and then were mounted with Prolong Gold antifade agent (Invitrogen). Fluorescence-labeled cells were observed under the confocal fluorescence microscope (LSM 510 META; Carl Zeiss GmbH, Zena, Germany).

Electrophoretic Mobility Shift Assay. The binding of activated TCF and the sequence of the TCF response element were evaluated using the TCF-electrophoretic mobility shift assay kit. Activated TCF was prepared as nuclear cell extracts from SW480 cells. Binding reactions containing 8 μ g of extracted protein and 1.5 pmol of biotin-labeled TCF binding probe were performed for 30 min at 15°C. The products were separated on 6% nondenaturing polyacrylamide gels and visualized using an enhanced chemiluminescence detection system.

Cell Invasion and Motility Assay. HCT116 cells (5×10^4 cells/chamber) were used for invasion and motility assays. The inner and outer surfaces on the filter membrane of a Transwell (Corning Life Sciences, Lowell, MA) were coated with 10 μ l of type I collagen (0.5 mg/ml) and 20 μ l of a 1:2 mixture of Matrigel-DMEM, respectively. Cells were plated on the upper chamber of the Matrigel-coated Transwell. The medium of the lower chambers also contained 0.1 mg/ml bovine serum albumin. The inserts were incubated for 24 h at 37°C. The cells that had invaded the outer surface of the membrane were fixed with methanol, stained with hematoxylin and eosin, and photographed.

To determine the cell motility, cells were seeded into the Transwell on the outer surface of the filter membrane coated with type I collagen. After incubation for 24 h, the membranes were fixed, stained, and photographed as described previously.

RNA Interference. RNA interference of β -catenin was performed using siRNA duplexes purchased from Bioneer (Daejeon, Korea). The sequence of the β -catenin siRNA was AUUACUAGAGCAGACA-GAUAGCACC and GGUGCUAUCUGUCUGCUCUAGUAAU for the sense and antisense strands, respectively. SW480 cells (2×10^5 cells/well in a six-well plate) were transfected with 10 nM concentrations of the siRNA duplexes using RNAiMAX (Invitrogen) according to the manufacturer's recommendations. Cells were also transfected with nonspecific control siRNA duplex for direct comparison. After 24 h of transfection, cells were collected, and the protein expression was determined using Western blotting.

In Vivo Tumor Xenograft Study. Female nude mice [5 weeks old, BALB/cA-nu (*nu/nu*)] were purchased from Central Laboratory Animal Inc. (Seoul, Korea) and acclimated for 1 week at $22 \pm 2^\circ\text{C}$ with a 12-h light/dark cycle in a pathogen-free environment. All animal experiments and care were conducted in a manner conforming to the Guidelines of the Animal Care and Use Committee of Ewha Womans University (permission number EWA2009-2-13). HCT116 cells were injected subcutaneously into the flanks of the mice (2×10^6 cells in 200 μ l of medium), and tumors were allowed to develop for 8 days until they reached approximately 80 mm³. The mice were randomized into vehicle control and treatment groups ($n = 5$). Magnolol (5 mg/kg b.wt.) was intraperitoneally injected in a solution containing 0.5% ethanol and 0.5% Cremophor (ethanol/Cremophor/H₂O, 0.5:0.5:99) in a volume of 200 μ l three times a week. The control group was treated with an equal volume of vehicle. Tumor volume was measured using calipers according to the following formula: tumor volume (mm³) = $3.14 \times L \times W \times H/6$, where L is the length, W is the width, and H is the height.

Statistical Analysis. Data are expressed as means \pm SD for the indicated number of independently performed experiments. Student's t test (SigmaStat 3.1; Systat Software, Inc. (San Jose, CA) was used for the determination of statistical significance. The difference was considered to be statistically significant when $P < 0.05$.

Results

Inhibitory Effects of Magnolol on β -Catenin/TCF Signaling. To search for small molecule inhibitors of β -catenin/TCF signaling, a TOPflash assay was used. In our evaluation of approximately 500 natural compounds, magnolol was identified as one of the most potentially active compounds in this assay system (Fig. 1A). Magnolol (Fig. 1A) exhibited potent inhibitory activity against the TOPflash reporter gene in a concentration-dependent manner (Fig. 1B). However, magnolol did not affect FOPflash activity (a mutant of the TCF binding site), suggesting that the inhibitory activity by magnolol is dependent on β -catenin/TCF signaling. To further elucidate whether the inhibitory activity of β -catenin/TCF signaling by magnolol is associated with target gene expression, downstream target genes, such as cyclin D1 or *c-myc*, were investigated with reverse transcriptase-PCR or real-time PCR analysis. Under normal conditions, the β -catenin level in HEK293 cells is relatively low, and, thus, the expression of target genes is maintained at a low level. However, the introduction of the β -catenin plasmid remarkably increased the expression of cyclin D1 and *c-myc* genes, target genes of β -catenin/TCF signaling. The elevated expression of the target genes by β -catenin was alleviated upon treatment with magnolol (Fig. 2A). Real-time PCR analysis also showed the down-regulation of cyclin D1 and *c-myc* gene expressions after treatment with magnolol in a concentration-dependent manner (Fig. 2B). In addition, target protein expression, specifically, the expression of the cyclin D1 and c-Myc proteins, was investigated in β -catenin-overexpressing cells. HEK293 cells were transfected with the β -catenin plasmid (1.5 μ g) for 24 h and then treated with magnolol for an additional 24 h. As shown in Fig. 2C, magnolol suppressed the expression of the cyclin D1 and c-Myc proteins. These results suggest that magnolol suppressed the expression of cyclin D1 and c-Myc through the inhibition of the β -catenin/TCF signaling pathway.

Inhibitory Effects of Magnolol on Wnt/ β -Catenin Signaling in HEK293-hFz1 Cells. Under normal conditions, APC/axin/GSK3 destruction complexes phosphorylate β -catenin, and the phosphorylated β -catenin is recognized by β -TrCP and then subsequently degraded by the ubiquitin-proteasome pathway. As a result, cytosolic β -catenin levels are maintained at a low level. In the presence of Wnt, the destruction complex is dissociated and cytosolic β -catenin is translocated into the nucleus to activate the target genes. To confirm the responsiveness of Wnt/ β -catenin signaling, various concentrations of Wnt3a-CM, which were conditioned from Wnt3a-producing L cells, were treated, and the TOPflash reporter gene assay was performed. Treatment with 50 and 100% Wnt3a-CM increased luciferase activity approximately 3-fold compared with that of the nontreated control (Supplemental Fig. 1A). On the basis of these findings, we further established that HEK293-hFz1 cells, which were anchored with the Wnt receptor human frizzled-1, enhance the responsiveness of Wnt signaling. Wnt3a-CM remarkably increased the TOPflash activity in HEK293-hFz1 cells compared with that in HEK293 cells (Supplemental Fig. 1B). The activation of Wnt-mediated signaling in HEK293-hFz1 cells was also confirmed by a significant increase in β -catenin expression in the cytoplasm and nucleus (Supplemental Fig. 1C). To evaluate whether magnolol suppresses Wnt/ β -catenin-mediated

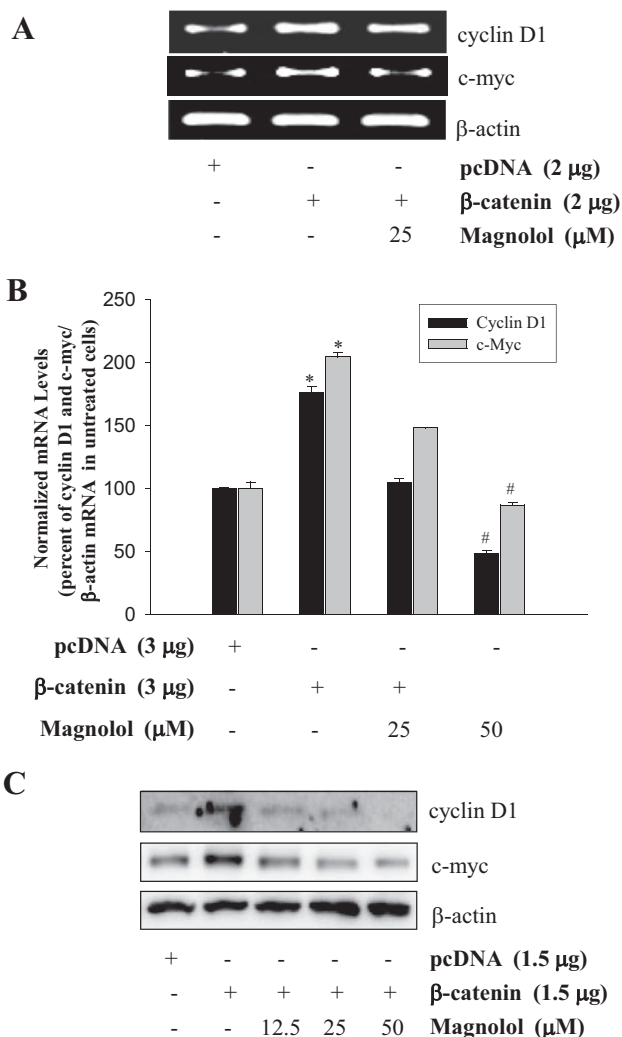


Fig. 2. Effect of magnolol on target gene expressions of β -catenin/TCF signaling. **A**, HEK293 cells were transfected with 2 μ g of either pcDNA or β -catenin plasmid, and then cells were treated with 25 μ M magnolol for 12 h. Total RNA was extracted and reverse transcribed with AMV reverse transcriptase and the oligo(dT)₁₅ primer. PCR was performed as described under *Materials and Methods*. The PCR products were separated by gel electrophoresis. The gel was stained with a SYBR Gold staining solution and visualized under a UV transilluminator. β -Actin gene served as an internal standard. **B**, inhibitory effects of magnolol on the cyclin D1 and *c-myc* expressions elevated by β -catenin in HEK293 cells. HEK293 cells were transfected with 3 μ g of either pcDNA or β -catenin plasmid and then treated with indicated concentrations (25 and 50 μ M) of magnolol for 12 h. The expression levels of cyclin D1 and *c-myc* were examined using real-time PCR. *, $P < 0.05$, compared with pcDNA alone-transfected cells; # $P < 0.05$, compared with the DMSO control. **C**, inhibitory effects of magnolol on the target protein expressions of β -catenin/TCF signaling in HEK293 cells. Cells were transfected with 1.5 μ g of either pcDNA or β -catenin plasmid. β -Actin gene was used as an internal standard. Data are representative of three independent experiments.

transcriptional activity in HEK293-hFz1 cells, the TOPflash activity and target gene expression were determined in Wnt3a-CM-treated HEK293-hFz1 cells. As shown in Fig. 3A, Wnt3a-CM markedly increased TOPflash activity, but magnolol abrogated the reporter gene activity in a concentration-dependent manner. Western blot analysis also demonstrated the suppression of the expression of target gene cyclin D1 by magnolol in Wnt3a-CM-treated HEK293-hFz1 cells (Fig. 3B).

To further elucidate the effect of magnolol on the expres-

sion level of Wnt-induced β -catenin, HEK293-hFz1 cells were treated with Wnt3a-CM in the presence or absence of magnolol for 24 h. Wnt3a-CM enhanced the expression of β -catenin in the cytosol and nucleus, and magnolol suppressed the Wnt3a-CM-induced expression of β -catenin in these cells (Fig. 3C). Because the increase in the degradation of β -catenin in the cytosolic fraction was not found by magnolol, the suppression of β -catenin in the nucleus fraction might be mainly associated with the inhibition of the nuclear translocation of β -catenin by magnolol. Immunofluorescence analysis also revealed the down-regulation of β -catenin by magnolol in the Wnt3a-CM-induced HEK293-hFz1 cells (Fig. 3D).

Inhibitory Effects of Magnolol on the β -Catenin/Tcf Signaling Pathway in Colorectal Cancer Cells. On the basis of the findings that magnolol suppressed Wnt3a-induced β -catenin and its target gene expression in HEK293-hFz1 cells, we further explored the effects of magnolol on β -catenin/TCF signaling in colon cancer cells. Colon cancer cells frequently possess mutations in APC or β -catenin and activated Wnt signaling as a result of the cellular accumulation of β -catenin. To investigate the inhibitory effect of magnolol on the transcriptional activity of β -catenin/TCF in colon cancer cells, the TOPflash reporter gene assay was performed in SW480 (truncated mutation of APC and wild-type β -catenin) or HCT116 (Ser45 deletion mutation of β -catenin and wild-type APC) human colon cancer cells. As shown in Fig. 4A, cells transfected with the TOPflash reporter showed the highest transcriptional activity, but magnolol suppressed the luciferase activity in human colon cancer cells.

To further clarify whether the suppressive effect of magnolol on the transcriptional activity of β -catenin/TCF is associated with the down-regulation of β -catenin, β -catenin levels in the cytosol and nucleus were examined. When treated with magnolol for 24 h, the expression of β -catenin in the cytosol and nucleus in SW480 cells was suppressed (Fig. 4B). Because magnolol decreased the expression of β -catenin in the nucleus of SW480 cells, it was assumed that the suppressive effect of magnolol on the transcriptional activity of β -catenin/TCF might be correlated with the decrease in the binding of β -catenin/TCF complexes to the promoter region of DNA. Therefore, the change in DNA-TCF complex binding was investigated using an electrophoretic mobility shift assay. Magnolol inhibited the binding of DNA and TCF through the TCF binding site (Fig. 4C). These findings suggest that the suppressive effects of magnolol on β -catenin/TCF transcription are associated with the down-regulation of β -catenin in the cytosol and nucleus and consequently the inhibition of binding between DNA and β -catenin/TCF complexes.

Inhibitory Effect of Magnolol on Target Gene Expressions and Cell Invasion and Migration Activity in Colon Cancer Cells. To assess the inhibitory effects of magnolol on β -catenin/TCF signaling in colon cancer cells, real-time PCR was used to determine the expression of β -catenin/TCF target genes. Cyclin D1 and *c-myc* genes are established target genes of the β -catenin-dependent pathway and are implicated in enhancement of cell proliferation (He et al., 1998; Liu et al., 2001).

As shown in Fig. 5A, treatment with magnolol suppressed the mRNA expression of cyclin D1 and *c-myc*. Further experiments showed that the expression of matrix MMP-7 and uPA, which are associated with cancer cell invasiveness and

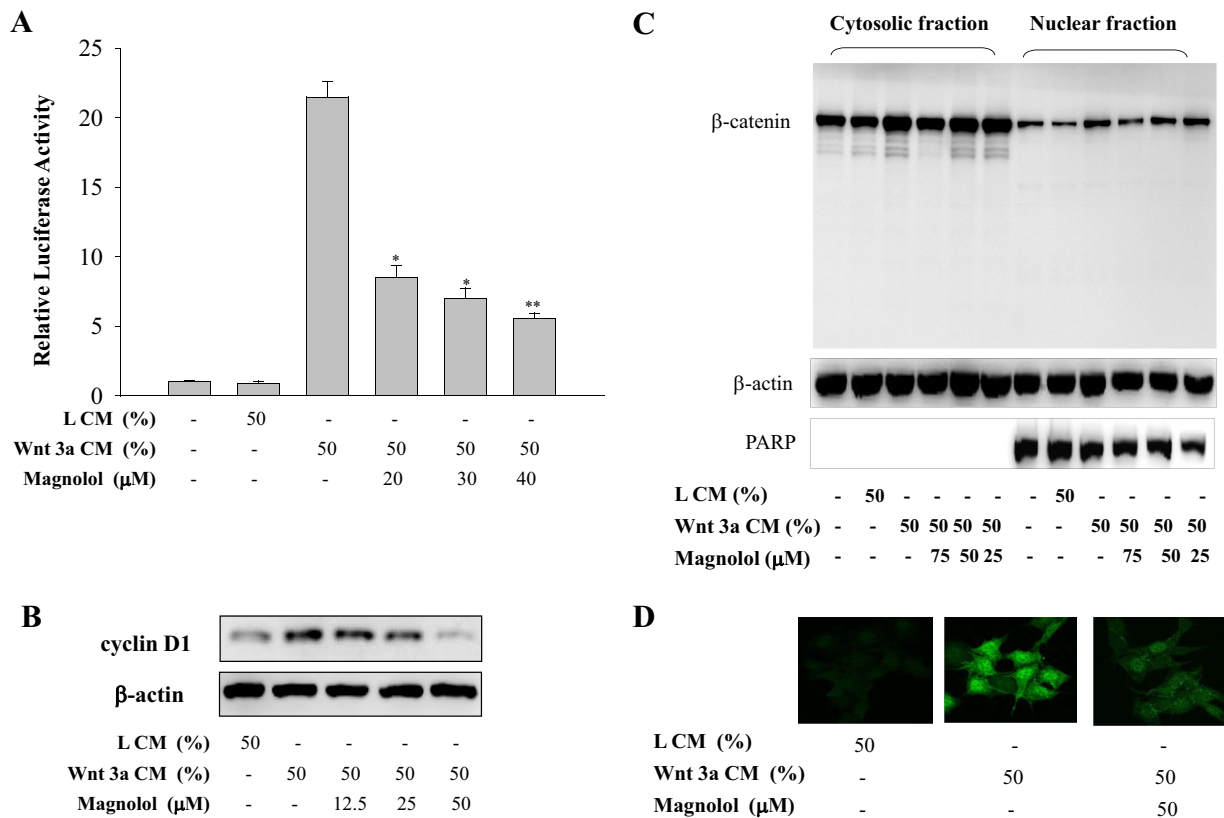


Fig. 3. Effects of magnolol on Wnt3a-induced β -catenin/TCF signaling in HEK293-hFz1 cells. **A**, HEK293-hFz1 cells were treated with various concentrations of magnolol (20, 30, and 40 μ M) in the presence of Wnt3a-CM. After a 24-h incubation, TOPflash activity was determined. *, $P < 0.05$; **, $P < 0.01$, compared with the DMSO control. **B**, inhibition of cyclin D1 expression by magnolol in Wnt3a-CM-treated HEK293-hFz1 cells. HEK293-hFz1 cells were treated with the indicated concentrations of magnolol for 24 h. The protein expression of cyclin D1 was measured by Western blot analysis. β -Actin was used as an internal standard. **C**, suppressive effect of magnolol on the expression of β -catenin induced by treatment with Wnt3a-CM in HEK293-hFz1 cells. HEK293-hFz1 cells were treated with magnolol in the presence of Wnt3a-CM, and then the cytosolic and nuclear fractions of the cell lysates were separated and analyzed for expression of β -catenin by Western blot. Anti-PARP was used for a marker of nuclear fraction. **D**, suppressive effect of magnolol on the expression of β -catenin in Wnt3a-CM-stimulated HEK293-hFz1 cells. The cells were treated with magnolol for 24 h in the presence of Wnt3a-CM and then were incubated with the anti- β -catenin antibody. The expression of β -catenin was investigated under confocal fluorescence microscopy (original magnification, 600 \times).

also target genes of β -catenin/TCF transcription (Brabletz et al., 1999; Wong and Pignatelli, 2002), were also significantly suppressed by magnolol (Fig. 5B).

Magnolol also suppressed the protein expressions of cyclin D1 and *c-myc*, but the expression of GSK3 β was not altered. However, magnolol remarkably induced the expression of E-cadherin (Fig. 5C). In addition, magnolol dose dependently inhibited the cancer cell invasion and motility through Transwell (Fig. 5D).

To further confirm whether magnolol regulates cyclin D1 and *c-myc* through β -catenin/TCF signaling, the direct effect of β -catenin siRNA was determined. SW480 cells were transfected with either β -catenin-specific siRNA or scrambled siRNA for 24 h, and then cells were incubated for an additional 24 h in serum-free media. The cells were treated with magnolol for 3 h, and then Western blotting was performed with cell lysates. Transfection with β -catenin siRNA effectively suppressed the protein expression of β -catenin. The expression of cyclin D1 and *c-myc* was also decreased by β -catenin siRNA. The expression of *c-myc* was dramatically suppressed compared with that of cyclin D1. However, magnolol enhanced the suppression of cyclin D1 expression in cells transfected with β -catenin siRNA in a concentration-dependent manner (Fig. 6).

Antiproliferative and Antitumor Activity of Magnolol in Colon Cancer Cells. Because the up-regulation of β -catenin plays a critical role in the proliferation of colon cancer cells (Verma et al., 2003), we assumed that the suppression of magnolol on the transcription of β -catenin/TCF might lead to growth inhibition in colon cancer cells. The antiproliferative effect of magnolol was investigated using the WST-1 assay in SW480 and HCT116 cells. Magnolol exhibited antiproliferative activity in a time- and concentration-dependent manner (Fig. 7A).

In addition, the antitumor activity of magnolol in vivo was assessed in a nude mouse xenograft model. HCT116 cells were subcutaneously implanted into nude mice. When tumor size reached ~ 80 mm³, magnolol (5 mg/kg) was administered intraperitoneally three times per week for 20 days. Tumor volume in the control group was approximately 1200 mm³ on day 28 after the cancer cells were inoculated. Treatment with magnolol effectively suppressed tumor growth, and the inhibitory effect in the magnolol-treated group (5 mg/kg) was approximately 54.6% compared with that of the control group (Fig. 7B), but body weight change on treatment with magnolol was negligible (Fig. 7C), and no overt toxicity was found under these experimental conditions.

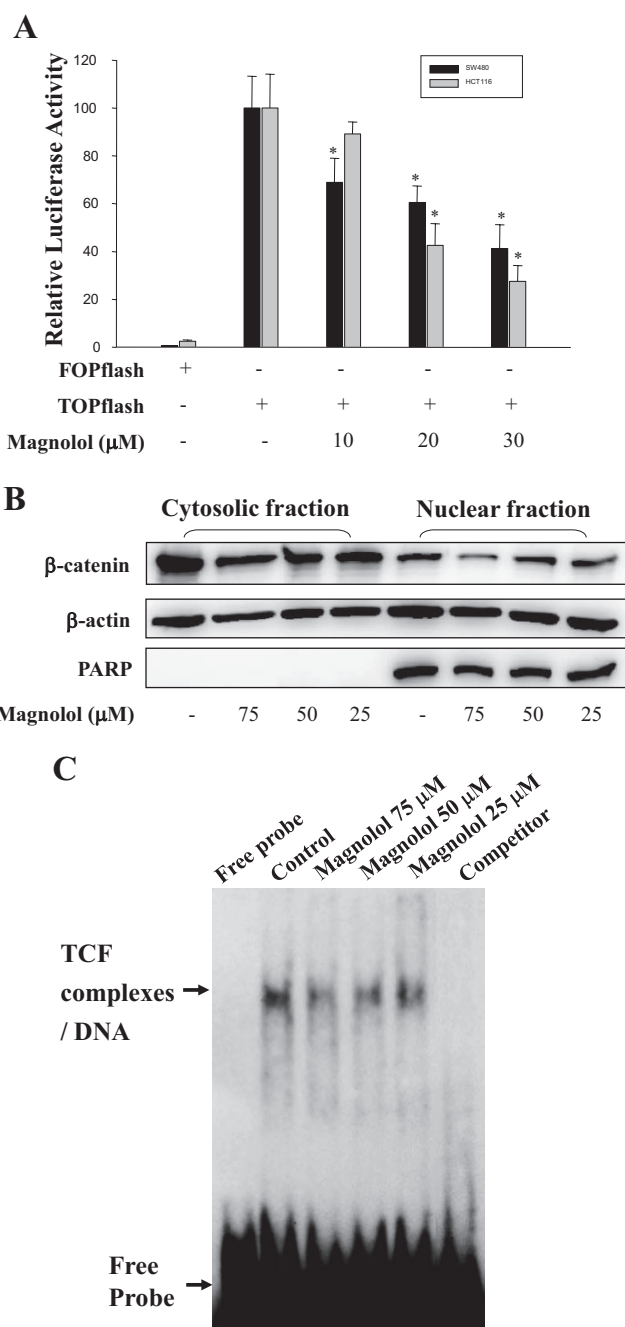


Fig. 4. Inhibitory effect of magnolol on the transcriptional activity of β -catenin/TCF in colon cancer cells. **A**, colon cancer (SW480 and HCT116) cells were cotransfected with TOPflash (or FOPflash) and *R. reniformis* for 24 h and then with magnolol for an additional 24 h. TOPflash or FOPflash activity was determined. *, $P < 0.05$, compared with the DMSO control. **B**, suppression of magnolol on the expression of β -catenin both in cytosolic and nuclear fractions in SW480 cells. SW480 cells were treated with magnolol for 24 h, and then the expression of β -catenin by was analyzed by Western blot. β -Actin was used as a loading control, and PARP was used as a nuclear fraction marker. **C**, inhibitory effects of magnolol on binding of TCF complexes to DNA. SW480 cells were treated with various concentrations of magnolol (25, 50, and 75 μ M) for 24 h, and then nuclear extracts were isolated. The electrophoretic mobility shift assay was performed with nuclear extracts from treated or untreated cells.

Discussion

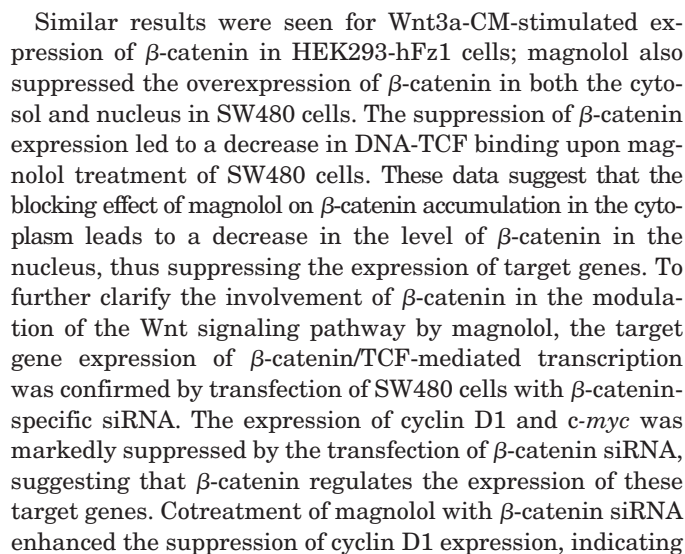
The Wnt/ β -catenin signaling pathway has been considered to play a crucial role in the early stages of carcinogenesis and the maintenance of colorectal cancers with either inactive

mutations of the APC tumor suppressor or active mutation of the β -catenin gene (Fearnhead et al., 2001; Liu et al., 2001; Bright-Thomas and Hargest, 2003). Indeed, high levels of β -catenin accumulate in colorectal cancer cells compared with the low levels of β -catenin seen in normal colonic cells (Polakis, 1997; Morin, 1999). Therefore, the modulation of Wnt/ β -catenin signaling is considered to be a promising drug target in the development of cancer chemotherapeutic agents. In our program to identify potential small molecule inhibitors of Wnt signaling, more than 500 compounds derived from natural products were evaluated using a cell-based TOPflash reporter gene assay in HEK293 cells. Several classes of natural products including curcuminoids, phenylpropanoids, and lignans exhibited potential inhibition of TOPflash activity. Among the active compounds, magnolol was one of the most prominent inhibitors of Wnt reporter gene activity. Therefore, in the present study, we examined the plausible mechanisms of action for the modulation of Wnt/ β -catenin signaling and the growth inhibition of human colorectal cancer cells by magnolol.

Previous studies have demonstrated the antiproliferation effect of magnolol on colon cancer cells by inhibition of DNA synthesis and activation of the Ras/Raf-1/extracellular signal-regulated kinase pathway (Lin et al., 2002; Hsu et al., 2007). However, the detailed mechanisms of the growth inhibition of magnolol in colorectal cancer cells remain to be elucidated. Furthermore, the modulation of Wnt/ β -catenin signaling by magnolol in colon cancer cells has not been identified.

Our cell-based reporter assays showed that magnolol effectively inhibited the Wnt/ β -catenin signaling pathway stimulated by β -catenin/TCF and Wnt3a-CM, suggesting that magnolol might target β -catenin or its downstream effectors. It was reported that the expression of cyclin D1 and *c-myc*, target genes of β -catenin/TCF transcription, was induced by the introduction of exogenous β -catenin cDNA in HEK293 cells (He et al., 1998; Shtutman et al., 1999; Tetsu and McCormick, 1999). To confirm whether magnolol affects the activation of Wnt signaling mediated by β -catenin overexpression, HEK293 cells were transfected with β -catenin cDNA, and the subsequent expression of cyclin D1 and *c-myc* was estimated by real-time PCR and reverse transcription PCR. The elevated expression of cyclin D1 and *c-myc* genes mediated by β -catenin was suppressed by magnolol, indicating that magnolol might inhibit β -catenin/TCF-mediated target gene expressions. Using hFz1-encoded established cells HEK293-hFz1, we also confirmed direct modulation of the Wnt signaling pathway by magnolol. Magnolol potentially suppressed Wnt-mediated downstream signaling with the inhibition of TOPflash activity, down-regulation of cyclin D1 expression, a decrease in β -catenin expression, and translocation of β -catenin into the nucleus in Wnt3a-CM-treated HEK293-hFz1 cells.

On the basis of our results for the effects of magnolol on β -catenin/TCF and Wnt3a-mediated signaling in the HEK293 and HEK293-hFz1 cell systems, we further evaluated the effects of magnolol on Wnt/ β -catenin signaling in human colorectal cancer cells, which possess intrinsically activated Wnt signaling through mutations in the APC or β -catenin genes as well as increased levels of both cytosolic and nuclear β -catenin (Weinberg, 2005; Sievers et al., 2006). The present study showed that magnolol inhibited the en-



dogenously activated transcriptional activity of β -catenin/TCF in both SW480 cells (APC mutants) and HCT116 cells (β -catenin mutants). The inhibition of β -catenin-mediated transcription by magnolol was also confirmed by the suppression of β -catenin/TCF target gene expressions, including the

expression of MMP-7, uPA, cyclin D1, and *c-myc* in colorectal cancer cells. These results suggest that magnolol suppresses cancer cell invasiveness and proliferation in the Wnt-activated colorectal cancer cells.

Similar results were seen for Wnt3a-CM-stimulated expression of β -catenin in HEK293-hFz1 cells; magnolol also suppressed the overexpression of β -catenin in both the cytosol and nucleus in SW480 cells. The suppression of β -catenin expression led to a decrease in DNA-TCF binding upon magnolol treatment of SW480 cells. These data suggest that the blocking effect of magnolol on β -catenin accumulation in the cytoplasm leads to a decrease in the level of β -catenin in the nucleus, thus suppressing the expression of target genes. To further clarify the involvement of β -catenin in the modulation of the Wnt signaling pathway by magnolol, the target gene expression of β -catenin/TCF-mediated transcription was confirmed by transfection of SW480 cells with β -catenin-specific siRNA. The expression of cyclin D1 and *c-myc* was markedly suppressed by the transfection of β -catenin siRNA, suggesting that β -catenin regulates the expression of these target genes. Cotreatment of magnolol with β -catenin siRNA enhanced the suppression of cyclin D1 expression, indicating

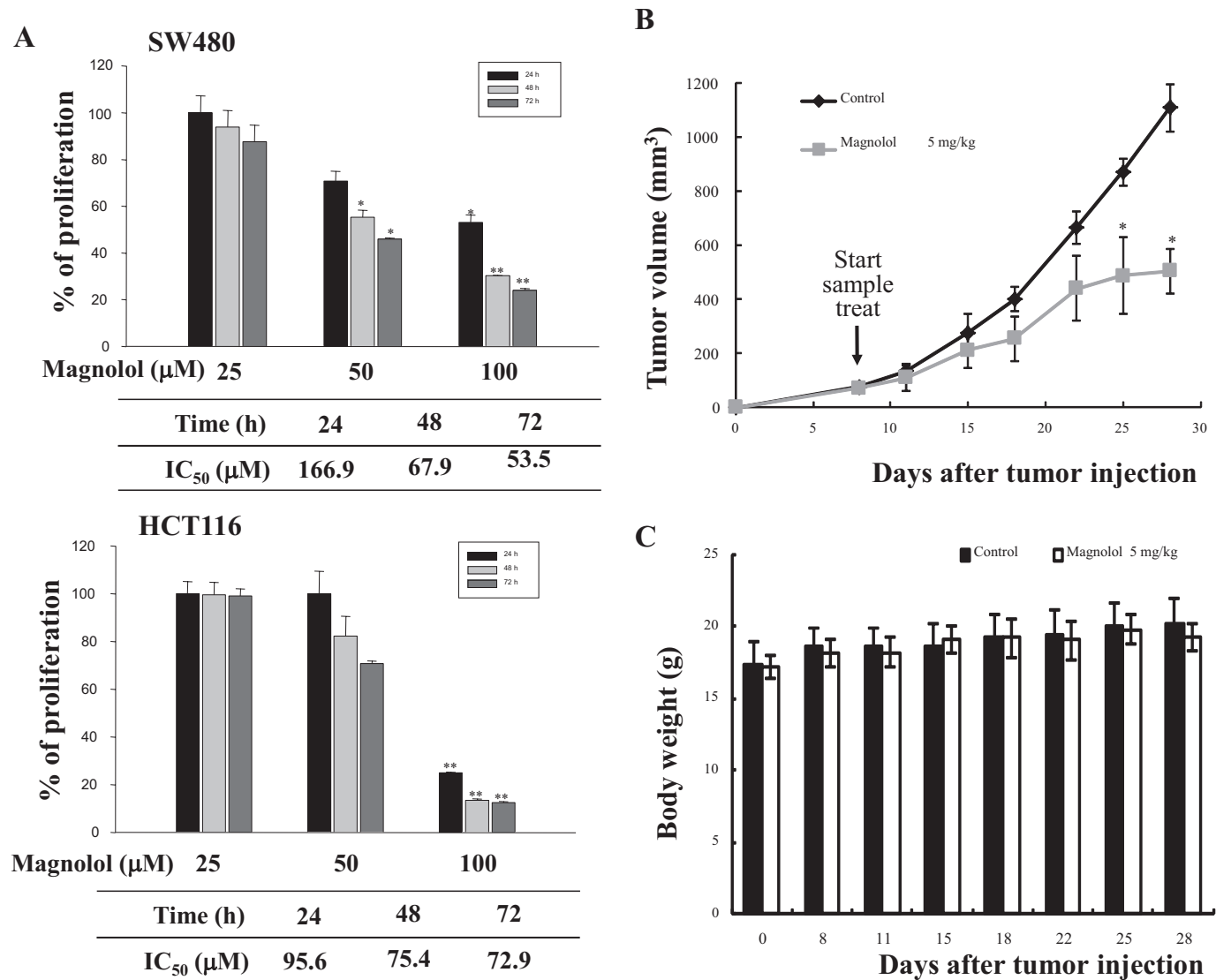


Fig. 7. Antiproliferation of magnolol in human colon cancer cells. **A**, SW480 or HCT116 colon cancer cells were treated with various concentrations of magnolol for 24, 48, or 72 h. Cell proliferation was measured using the WST-1 assay. Percentage of proliferation was determined by comparison with vehicle-treated control cells ($n = 3$). *, $P < 0.05$; **, $P < 0.01$, compared with the DMSO control. **B** and **C**, inhibition of tumor growth by magnolol (**B**) and body weight change in the HCT116 xenograft nude mouse model (**C**). HCT116 cells (2×10^6 cells) were injected subcutaneously into the flanks of nude mice, and tumors were allowed to develop for 8 days until they reached $\sim 80 \text{ mm}^3$; then treatment was initiated ($n = 5$). The mice were administered magnolol (5 mg/kg i.p.) three times a week in a solution containing 0.5% ethanol and 0.5% Cremophor (ethanol/Cremophor/ H_2O , 0.5:0.5:99) in a volume of $200 \mu\text{l}$ for 20 days. The control group was treated with an equal volume of vehicle. Tumor volume and body weight were monitored every 2 to 3 days. *, $P < 0.05$, compared with the vehicle-alone injection.

the involvement of β -catenin in the modulation of the Wnt signaling pathway by magnolol.

To further elucidate the regulation of β -catenin by magnolol, we determined the expression of one plausible modulator, E-cadherin. E-cadherin is a transmembrane glycoprotein that mediates tight adhesion between cells, and the cytoplasmic domain of E-cadherin interacts with α -, β -, and γ -catenin. In a previous study, a signal to the nucleus via β -catenin was hypothesized to be controlled by E-cadherin because of the dual roles of β -catenin: induction of the nuclear signal and cadherin-mediated adhesion at the plasma membrane (Cox et al., 1996; Gottardi et al., 2001). In the present study, we found that magnolol enhanced the expression of E-cadherin in Wnt3a-CM-stimulated HEK293-hFz1 cells, and the level of E-cadherin was also increased by magnolol in SW480 and HCT116 cells. These data suggest that

magnolol might increase the expression of E-cadherin, and, thus, the level of cytosolic β -catenin should be down-regulated. The up-regulation of E-cadherin and suppression of MMP-7 and uPA by magnolol were also somewhat correlated with the inhibition of colon cancer cell invasion and motility. However, further studies are needed to elucidate whether the magnolol-induced E-cadherin binds and sequesters β -catenin in the cytosol. Recent reports by Thorne et al. (2010) show that the inhibition of Wnt signaling by the small molecule inhibitor pyrvinium is associated with the activation of casein kinase 1 α , a component of the β -catenin destruction complex in the cytosol. Thus, the precise mechanisms of the upstream regulation of β -catenin in the cytosol by magnolol still remain to be clarified.

Because magnolol suppressed the expressions of cyclin D1 and *c-myc*, major factors in cancer cell proliferation, further

experiments were designed to investigate whether magnolol affects the proliferation of cancer cells by altering Wnt signaling in human colorectal cancer cells. Magnolol effectively inhibited the proliferation of both SW480 and HCT116 cells in a concentration-dependent manner in vitro. Tumor growth was also suppressed by magnolol in a nude mouse xenograft model bearing HCT116 cells (Fig. 7). These data suggest that the antiproliferative effects of magnolol in colorectal cancer cells might partly be associated with the down-regulation of Wnt signaling.

In summary, the present study demonstrates that magnolol might be a potential candidate in the development of small molecule inhibitors of Wnt signaling. Therefore, magnolol might be useful for treating sporadic colon cancer cells either alone or in combination with other chemotherapeutic agents.

Authorship Contributions

Participated in research design: Kang, H. J. Park, Chung, Min, E. J. Park, and S. K. Lee.

Conducted experiments: Kang, H. J. Park, Chung, M. A. Lee, and Shin.

Contributed new reagents or analytic tools: Kang, H. J. Park, and Chung.

Performed data analysis: Kang, H. J. Park, Chung, and S. K. Lee.

Wrote or contributed to the writing of the manuscript: Kang, H. J. Park, Chung, and S. K. Lee.

References

- Battle TE, Arbisser J, and Frank DA (2005) The natural product honokiol induces caspase-dependent apoptosis in B-cell chronic lymphocytic leukemia (B-CLL) cells. *Blood* **106**:690–697.
- Bienzi M (2002) The subcellular destinations of APC proteins. *Nat Rev Mol Cell Biol* **3**:328–338.
- Brabletz T, Jung A, Dag S, Hlubek F, and Kirchner T (1999) β -Catenin regulates the expression of the matrix metalloproteinase-7 in human colorectal cancer. *Am J Pathol* **155**:1033–1038.
- Bradford MM (1976) A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal Biochem* **72**:248–254.
- Bright-Thomas RM and Hargest R (2003) APC, β -catenin and hTCF-4; an unholy trinity in the genesis of colorectal cancer. *Eur J Surg Oncol* **29**:107–117.
- Clark AM, El-Ferali FS, and Li WS (1981) Antimicrobial activity of phenolic constituents of *Magnolia grandiflora* L. *J Pharm Sci* **70**:951–952.
- Cong F and Varmus H (2004) Nuclear-cytoplasmic shuttling of Axin regulates subcellular localization of β -catenin. *Proc Natl Acad Sci USA* **101**:2882–2887.
- Cox RT, Kirkpatrick C, and Peifer M (1996) Armadillo is required for adherens junction assembly, cell polarity, and morphogenesis during *Drosophila* embryogenesis. *J Cell Biol* **134**:133–148.
- Fearnhead NS, Britton MP, and Bodmer WF (2001) The ABC of APC. *Hum Mol Genet* **10**:721–733.
- Gottardi CJ, Wong E, and Gumbiner BM (2001) E-cadherin suppresses cellular transformation by inhibiting β -catenin signaling in an adhesion-independent manner. *J Cell Biol* **153**:1049–1060.
- He TC, Sparks AB, Rago C, Hermeking H, Zawel L, da Costa LT, Morin PJ, Vogelstein B, and Kinzler KW (1998) Identification of c-MYC as a target of the APC pathway. *Science* **281**:1509–1512.
- Hsu YF, Lee TS, Lin SY, Hsu SP, Juan SH, Hsu YH, Zhong WB, and Lee WS (2007) Involvement of Ras/Raf-1/ERK actions in the magnolol-induced upregulation of p21 and cell-cycle arrest in colon cancer cells. *Mol Carcinog* **46**:275–283.
- Huang SH, Chen Y, Tung PY, Wu JC, Chen KH, Wu JM, and Wang SM (2007) Mechanisms for the magnolol-induced cell death of CGTH W-2 thyroid carcinoma cells. *J Cell Biochem* **101**:1011–1022.
- Huelsken J and Behrens J (2002) The Wnt signalling pathway. *J Cell Sci* **115**:3977–3978.
- Ikeda S, Kishida S, Yamamoto H, Murai H, Koyama S, and Kikuchi A (1998) Axin, a negative regulator of the Wnt signaling pathway, forms a complex with GSK-3 β and β -catenin and promotes GSK-3 β -dependent phosphorylation of β -catenin. *EMBO J* **17**:1371–1384.
- Ishitsuka K, Hideshima T, Hamasaki M, Raje N, Kumar S, Hideshima H, Shiraishi N, Yasui H, Roccaro AM, Richardson P, et al. (2005) Honokiol overcomes conventional drug resistance in human multiple myeloma by induction of caspase-dependent and -independent apoptosis. *Blood* **106**:1794–1800.
- Ko CH, Chen HH, Lin YR, and Chan MH (2003) Inhibition of smooth muscle contraction by magnolol and honokiol in porcine trachea. *Planta Med* **69**:532–536.
- Kong ZL, Tzeng SC, and Liu YC (2005) Cytotoxic neolignans: an SAR study. *Bioorg Med Chem Lett* **15**:163–166.
- Lee SY, Yuk DY, Song HS, Yoon do Y, Jung JK, Moon DC, Lee BS, and Hong JT (2008) Growth inhibitory effects of obovatol through induction of apoptotic cell death in prostate and colon cancer by blocking of NF- κ B. *Eur J Pharmacol* **582**:17–25.
- Lin SY, Liu JD, Chang HC, Yeh SD, Lin CH, and Lee WS (2002) Magnolol suppresses proliferation of cultured human colon and liver cancer cells by inhibiting DNA synthesis and activating apoptosis. *J Cell Biochem* **84**:532–544.
- Liu J, Stevens J, Rote CA, Yost HJ, Hu Y, Neufeld KL, White RL, and Matsunami N (2001) Siah-1 mediates a novel β -catenin degradation pathway linking p53 to the adenomatous polyposis coli protein. *Mol Cell* **7**:927–936.
- Liu X, Rubin JS, and Kimmel AR (2005) Rapid, Wnt-induced changes in GSK3 β associations that regulate β -catenin stabilization are mediated by G α proteins. *Curr Biol* **15**:1989–1997.
- Morin PJ (1999) β -Catenin signaling and cancer. *Bioessays* **21**:1021–1030.
- Phelps RA, Broadbent TJ, Stafforini DM, and Jones DA (2009) New perspectives on APC control of cell fate and proliferation in colorectal cancer. *Cell Cycle* **8**:2549–2556.
- Polakis P (1997) The adenomatous polyposis coli (APC) tumor suppressor. *Biochim Biophys Acta* **1332**:F127–F147.
- Shan BE, Wang MX, and Li RQ (2009) Quercetin inhibit human SW480 colon cancer growth in association with inhibition of cyclin D1 and survivin expression through Wnt/ β -catenin signaling pathway. *Cancer Invest* **27**:604–612.
- Shtutman M, Zhurinsky J, Simcha I, Albanese C, D'Amico M, Pestell R, and Ben-Ze'ev A (1999) The cyclin D1 gene is a target of the β -catenin/LEF-1 pathway. *Proc Natl Acad Sci USA* **96**:5522–5527.
- Sievers S, Fritzsche C, Grzegorzczak M, Kuhnen C, and Müller O (2006) Absolute β -catenin concentrations in Wnt pathway-stimulated and non-stimulated cells. *Biomarkers* **11**:270–278.
- Teng CM, Chen CC, Ko FN, Lee LG, Huang TF, Chen YP, and Hsu HY (1988) Two antiplatelet agents from *Magnolia officinalis*. *Thromb Res* **50**:757–765.
- Tetsu O and McCormick F (1999) β -Catenin regulates expression of cyclin D1 in colon carcinoma cells. *Nature* **398**:422–426.
- Thorne CA, Hanson AJ, Schneider J, Tahinci E, Orton D, Cselenyi CS, Jernigan KK, Meyers KC, Hang BI, Waterson AG, et al. (2010) Small-molecule inhibition of Wnt signaling through activation of casein kinase 1 α . *Nat Chem Biol* **6**:829–836.
- Verma UN, Surabhi RM, Schmaltieg A, Becerra C, and Gaynor RB (2003) Small interfering RNAs directed against β -catenin inhibit the in vitro and in vivo growth of colon cancer cells. *Clin Cancer Res* **9**:1291–1300.
- Weinberg RA (2005) Multistep tumorigenesis, in *The Biology of Cancer* (Weinberg RA ed), pp 399–462, Garland Science, New York.
- Wong NA and Pignatelli M (2002) β -Catenin—a linchpin in colorectal carcinogenesis? *Am J Pathol* **160**:389–401.
- Zhong WB, Wang CY, Ho KJ, Lu FJ, Chang TC, and Lee WS (2003) Magnolol induces apoptosis in human leukemia cells via cytochrome c release and caspase activation. *Anticancer Drugs* **14**:211–217.

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